### Organic & Biomolecular **Chemistry**

Cite this: Org. Biomol. Chem., 2011, **9**, 2606

# www.rsc.org/obc **COMMUNICATION**

## **Novel hemicyanine dye as colorimetric and fluorometric dual-modal chemosensor for mercury in water†**

**Yuanyuan Li, Song He, Yan Lu\* and Xianshun Zeng\***

*Received 22nd November 2010, Accepted 7th February 2011* **DOI: 10.1039/c0ob01060j**

**A novel water soluble Hg2+-selective chemosensor 1 with** hemicyanine as fluorescent reporting group and  $NO<sub>2</sub>Se<sub>2</sub>$ **chelating unit as ion binding site was reported. Chemosensor 1 shows a specific Hg2+ selectivity and discrimination between Hg2+ and chemically similar ions in conjunction with a visible colorimetric change from red to colorless, potentially leading to both "naked-eye" and fluorometric detection of Hg2+ cations.**

The development of highly selective chemosensors for metal ions is particularly important, since some metal ions can have extremely harmful effects on humans and the environment.**<sup>1</sup>** Mercury is a particular concern worldwide because of its high toxicity. Mercury species are considered as prevalent toxic pollutants in the environment because both elemental and ionic mercury can be converted by bacteria in the environment to methylmercury, which subsequently bioaccumulates through the food chain.<sup>2</sup> When absorbed in the human body, mercury causes a wide variety of diseases such as prenatal brain damage, serious cognitive and motion disorders**2b,3** through damage to the central nervous system, DNA, mitosis, and the endocrine system.**<sup>4</sup>** Thus, concerns over toxic damage from mercury provide the motivation to explore the selective monitoring of  $Hg^{2+}$  in biological and environmental samples. Since chromo- or fluoroionophores are proven to be highly effective for the determination in terms of ease of handling and simplicity of equipment, much effort has been paid to the development of optical chemosensors that selectively respond to the mercuric ion.**<sup>5</sup>** To date, many chemosensors for mercury ion based on photoinduced electron transfer (PET), intramolecular charge transfer (ICT), and fluorescence resonance energy transfer (FRET) signaling mechanisms have been developed.**<sup>5</sup>** Dosimeters**<sup>6</sup>** and allosteric interaction sensors**<sup>7</sup>** have also been reported. Significantly, the molecular design of multimodal chemosensors for mercury is still a challenging but extremely attractive method because they have important features that permit multimodal

*Key Laboratory of Display Materials & Photoelectric Devices, Ministry of Education, School of Materials Science & Engineering, Tianjin University of Technology, Tianjin, 300384, China. E-mail: luyan@tjut.edu.cn, xshzeng@tjut.edu.cn; Fax: +86-22-60215226; Tel: +86-22-60216748*

signal detection when receptors interact with analytes; a multimodal signal system can minimize the measurement errors because of factors such as phototransformation, receptor concentrations, and environmental effects. In particular, some of them can be easily read with the naked eye. Consequently, the development of multimodal and naked eye chemosensors for  $Hg^{2+}$  has the important feature that they can be used to evaluate the analytes rapidly. In particular, most of the reported colorimetric sensors for Hg2+ are used in mixed solvents (water/organic solvent);**6g,8** only a few of them are used in neat aqueous solution.**5j,9**

In this regard, we are interested in hemicyanine-based dyes, known for their excellent spectroscopic properties of large molar extinction coefficient ( $\varepsilon \approx 10^4$  M<sup>-1</sup> cm<sup>-1</sup> in the visible region) and good fluorescence quantum yield, and which have found applications in many fields, such as the study of complex biological systems as molecular probes, photosensitizers in dye-sensitized solar cells *etc*, but few as chemosensors.**<sup>10</sup>** Taking into account the possibility of using positively charged hemicyanine dyes not only for colorimetric but also for fluorometric mercury sensing, increasing the water solubility and sensitivity by using fluorescence emission spectroscopic methods, we report our successful development of a novel hemicyanine-based water soluble colorimetric and fluorometric chemosensor 1 selective for Hg<sup>2+</sup> based on the mechanism of internal charge transfer (ICT).

We report herein a new hemicyanine-based water soluble chemosensor **1** (Scheme 1), which displayed high  $Hg^{2+}$  selectivity both in water and in ethanol–water solution. The chemosensor **1** was synthesized from **2** in two steps. As shown in Scheme 1, aldehyde **3** was prepared by the reaction of **2** with sodium salt



**Scheme 1** Structure and the synthesis of the chemosensor **1**.

<sup>†</sup> Electronic supplementary information (ESI) available: Experimental procedures and spectral data for all new compounds or other electronic format. See DOI: 10.1039/c0ob01060j

of 2-selenolethanol prepared *in situ* by the reaction of 1,2-(bis-2-hydroxylethyl)diselenide with NaBH4 in THF–ethanol mixture. Then, 1 was facilely obtained by the condensation of 1,2,3,3<sup>'</sup>tetramethylindolium iodide with **3** in 83% yield. The anion exchange was achieved by refluxing the ethanol solutions **1** with excess KBF4. The structure of **1** was confirmed by MS, NMR and elemental analysis. The synthesized chemosensor **1** composed of an ionophore for selective recognition of metal ions is constituted of an  $NSe<sub>2</sub>O<sub>2</sub>$ , chelating unit as cation binding site as well as a hemicyanine fluorophore unit which is responsible for signal transduction during spectroscopic studies.

Chemosensor 1 is soluble in  $H_2O$  or in a mixed solvent of  $H_2O$ –EtOH. The UV/Vis spectra of 1 both in  $H_2O$  and in aqueous ethanol show the characteristic intense charge-transfer band occurring in the long-wavelength region between 425 and 600 nm, which peaked at 529 nm in water ( $\varepsilon = 7.1 \times 10^4$  M<sup>-1</sup> cm<sup>-1</sup>) and 542 nm in EtOH-H<sub>2</sub>O (1:1, v/v) ( $\varepsilon = 8.3 \times 10^4$  M<sup>-1</sup> cm-<sup>1</sup> ) (Fig. 1a and 1b). It is interesting to note that the addition of 10 equiv of Hg2+ into the red solutions of **1** rapidly generated a colorless solution, while other ions, such as  $K^+$ ,  $Na^+$ ,  $NH_4^+$ ,  $Al^{3+}$ ,  $Co<sup>2+</sup>, Fe<sup>3+</sup>, Pb<sup>2+</sup>, Ag<sup>+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup>, Cd<sup>2+</sup>, Ni<sup>2+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup> gave no$ color change in either  $H_2O$  or in aqueous ethanol, which means that the two selenium atoms within **1** bound more efficiently to with Hg<sup>2+</sup>. This interesting feature revealed that 1 can serve as a selective "naked-eye" chemosensor for  $Hg^{2+}$  (Fig. 2).



Fig. 1 UV-vis responses of chemosensor  $1(11.5 \mu M)$  upon the addition of the nitrate salts (10.0 equiv) of Na<sup>+</sup>, K<sup>+</sup>, NH<sub>4</sub><sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Pb<sup>2+</sup>, Hg<sup>2+</sup>,  $Co^{2+}$ ,  $Cd^{2+}$ ,  $Zn^{2+}$ ,  $Ni^{2+}$ ,  $Cu^{2+}$ ,  $Cr^{3+}$ ,  $Fe^{3+}$  and  $Ag^+$  (a) in ethanol–H<sub>2</sub>O (1 : 1,  $v/v$ ) and (b) in  $H<sub>2</sub>O$ .

The metal ion binding properties and selectivity of chemosensor **1** were firstly investigated by absorbance changes both in  $H_2O$  and in aqueous ethanol (ethanol–water,  $1:1$ ,  $v/v$ ) solutions with use of metal nitrate salts. Fig. 1a displays the absorbance changes of



**Fig. 2** Photograph of 10  $\mu$ M **1** in the presence of different nitrate salts:  $(10.0 \text{ equiv})$  of Na<sup>+</sup>, K<sup>+</sup>, NH<sub>4</sub><sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Pb<sup>2+</sup>, Hg<sup>2+</sup>, Co<sup>2+</sup>, Cd<sup>2+</sup>, Zn<sup>2+</sup>,  $Ni^{2+}$ ,  $Cu^{2+}$ ,  $Cr^{3+}$ ,  $Fe^{3+}$  and  $Ag^{+}$  in  $H_2O$ .

**1** in the presence and absence of metal ions in EtOH–H<sub>2</sub>O (1:1,  $v/v$ ). As can be seen from Fig. 1a, the response of 1 to Hg<sup>2+</sup> leads to a variation in absorption. When 10 equiv of  $Hg^{2+}$  was added to the solution of  $1$  (11.5  $\mu$ M), the absorption peak at 542 nm diminished drastically, while a new absorption band appeared at  $400 \text{ nm}$  ( $\varepsilon = 2.0 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$ ). There was a large decrease (54fold) of absorbance at  $\lambda_{\text{max}} = 542 \text{ nm}$  and a mild enhancement (42-fold) of absorbance at  $\lambda_{\text{max}} = 400$  nm upon the addition of 10 equiv of Hg2+. Fig. 1b displays the absorbance changes of **1** in the presence of metal ions in water. When 10 equiv of  $Hg^{2+}$  was added to the solution of  $1(11.5 \mu M)$ , the absorption peak at 529 nm diminished drastically, while a new absorption band appeared at 384 nm ( $\varepsilon = 1.7 \times 10^4$  M<sup>-1</sup> cm<sup>-1</sup>). There was a large decrease (74-fold) of absorbance at  $\lambda_{\text{max}} = 529$  nm and a mild enhancement (27-fold) of absorbance at  $\lambda_{\text{max}} = 384$  nm upon the addition of 10 equiv of  $Hg^{2+}$  (Fig. 1b). As shown in Fig. 1b, under identical conditions to Hg<sup>2+</sup> ion, small increases in absorbance at  $\lambda_{\text{max}} = 529$ nm were observed in the UV/vis spectra of **1** upon addition of 10 equiv of K<sup>+</sup>, Na<sup>+</sup>, NH<sub>4</sub><sup>+</sup>, Al<sup>3+</sup>, Co<sup>2+</sup>, Fe<sup>3+</sup>, Pb<sup>2+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup>, Cd<sup>2+</sup>,  $Ni<sup>2+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup> but Ag<sup>+</sup> decreases the absorption to some extent$ in  $H_2O$ . From these results, it can be considered that the decrease of absorption intensity over 500 nm and the increase of absorption intensity around 400 nm caused by  $Hg<sup>2+</sup>$  addition are due to the change in the photophysical properties of the chromophore itself, and arise *via* the ICT mechanism.

The absorbance changes of  $Hg^{2+}$  titration of 1 were investigated both in H<sub>2</sub>O and in aqueous ethanol (ethanol–water, 1:1,  $v/v$ ) solutions. As shown in Fig. 3, upon addition of  $Hg^{2+}$  to the solution of **1**, the absorbance at 542 nm in aqueous ethanol and 529 nm in water decreased sharply, while that at 412 nm in aqueous ethanol and 404 nm in water increased significantly, which induced a color change from red to colorless. The isosbestic points at 471 nm (in EtOH–H<sub>2</sub>O, 1:1, v/v) and 457 nm (in H<sub>2</sub>O) were observed, indicating the formation of a new complex between 1 and  $Hg^{2+}$ . A linear dependence of the ratio of absorbance as a function of  $Hg^{2+}$ concentration both in water and in aqueous ethanol was observed.

Next, the nitrate salts of  $K^+$ , Na<sup>+</sup>, NH<sub>4</sub><sup>+</sup>, Al<sup>3+</sup>, Co<sup>2+</sup>, Fe<sup>3+</sup>, Pb<sup>2+</sup>, Ag<sup>+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup>, Cd<sup>2+</sup>, Ni<sup>2+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup> and Hg<sup>2+</sup> ions were used to evaluate the metal ion binding properties and the selectivity of chemosensor  $1(1 \mu M)$  by means of fluorescence spectra in EtOH–H<sub>2</sub>O (1 : 1, v/v) and in H<sub>2</sub>O, respectively. Among the metal ions examined, **1** showed a selective fluorescence decrease only with Hg<sup>2+</sup> both in EtOH–H<sub>2</sub>O (1 : 1, v/v) and in H<sub>2</sub>O, indicating that 1 displayed a high  $Hg^{2+}$  selectivity (Fig. 4 and Fig. S1, ESI†). The fluorescence spectra were obtained by excitation of the hemicyanine fluorophore at 540 nm.

To achieve a reliable binding constant (*K*) *via* fluorescence titration,**<sup>11</sup>** the titrations were conducted in a very dilute solution  $(1.0 \times 10^{-6} \text{ M})$ . From the fluorescence titration experiments (Fig. 5), clear "on–off" fluorescence changes of 1 to Hg<sup>2+</sup> were observed.



**Fig. 3** Absorbance spectra of  $1(10 \mu M)(a)$  in EtOH–H<sub>2</sub>O (1:1, v/v) and (b) in  $H_2O$  in the presence of different amounts of  $Hg^{2+}$ . Inset: the ratio of absorbance at 560 nm and absorbance at 400 nm as a function of  $Hg^{2+}$ concentration.



**Fig. 4** Fluorescence spectra of  $1(1 \mu M)$  in ethanol–water  $(1:1, v/v)$  and in H<sub>2</sub>O with 10 equiv of K<sup>+</sup>, Na<sup>+</sup>, NH<sub>4</sub><sup>+</sup>, Al<sup>3+</sup>, Co<sup>2+</sup>, Fe<sup>3+</sup>, Pb<sup>2+</sup>, Ag<sup>+</sup>, Cu<sup>2+</sup>,  $Zn^{2+}$ ,  $Cd^{2+}$ ,  $Ni^{2+}$ ,  $Ca^{2+}$ ,  $Mg^{2+}$  and  $Hg^{2+}$ . For the entire test, excitation and emission were performed at 540 and 596 nm in ethanol–water  $(1:1, v/v)$ and at 520 and 591 nm in water.

When  $Hg^{2+}$  was added to the solution, a significant decrease of the fluorescence intensity of 596 nm (in aqueous ethanol)/587 nm (in water) and with a blue shift to 575 nm was observed. The quenching is found to follow a Benesi–Hildebrand equation and may be attributed to the nitrogen lone-pair electrons induced by the complexation of  $Hg^{2+}$ . A linear dependence of the ratio of fluorescence intensity  $[1][Hg^{2+}]/(F_0 - F_i)$  at 594 nm (in aqueous ethanol)/590 nm (in water) as a function of  $Hg^{2+}$  concentration and the data from Job's plots from emission spectra (Fig. S2, ESI†) assumed a 1 : 1 stoichiometry for the **1**–Hg2+ complex. The binding constant was found to be  $8.81 \times 10^4$  M<sup>-1</sup> in aqueous ethanol and  $4.30 \times 10^5$  M<sup>-1</sup> in water, respectively, indicating that the binding ability of **1** in water is much stronger than that in aqueous ethanol. From the changes in  $Hg^{2+}$ -dependent fluorescence intensity (Fig. S3, ESI), the detection limit was estimated to be  $5.0 \times 10^{-8}$  M,<sup>12a</sup> indicating that the limit of detection of  $1$  to  $Hg^{2+}$  met the discharge limit for industrial waste water according to the China SA standard,**12b** or the US EPA standard.**12c**

Ion interference experiments were carried out both in H<sub>2</sub>O and in aqueous ethanol (ethanol–water, 1 : 1, v/v) solvent systems for evaluation of the sensing ability of 1 toward  $Hg^{2+}$  in the presence of 50 equiv of a series of background metal ions, and the results are shown in Fig. 6. While  $Ag^+$  had 5% background quenching



**Fig. 5** Fluorescent titration spectra: (a)  $1(1 \mu M)$  in the presence of different concentrations of Hg<sup>2+</sup> in ethanol–water (1 : 1, v/v), [Hg<sup>2+</sup>] = 0, 1, 3, 5, 10, 15, 20, 25, 30, 40, 50, 60, 80, 100, 150, 200 equiv. Inset: Fluorescence intensity  $F_i$  as a function of Hg<sup>2+</sup> concentration.  $\lambda_{ex} = 540$  nm; (b) **1** (1  $\mu$ M) in the presence of different concentrations of Hg<sup>2+</sup> in water,  $[Hg^{2+}] = 0, 0.4$ , 0.8, 1, 1.5, 2, 3.5, 7, 9, 10, 20, 30, 35, 40, 50, 60, 70, 80, 90 equiv. Inset: the fluorescence at 590 nm of  $1(1 \mu M)$  as a function of the Hg<sup>2+</sup> concentration.  $\lambda_{\rm ex} = 530$  nm.



**Fig. 6** Quench ratio  $((I_0 - I)/I_0)$  of fluorescence intensity of 1 (1  $\mu$ M) in H<sub>2</sub>O and in EtOH–H<sub>2</sub>O (1 : 1, v/v) upon the addition of 10 equiv Hg<sup>2+</sup> in the presence of 50 equiv background metal ions.

in aqueous ethanol, other metal ions did not cause any effect. No interference was observed in  $H_2O$  in the presence of 50 equiv of a series of metal ions. On the other hand, almost no color change is observed in the presence of metal ions other than  $Hg^{2+}$ . When 10 equiv of  $Hg^{2+}$  is added to these solutions, the red colour disappears immediately and the spectra are almost identical to that obtained in the presence of  $Hg^{2+}$  alone.

For practical applicability, the most suitable pH conditions of chemosensor **1** were evaluated by means of absorption spectra. The absorption intensities at different pH values of the apo and  $Hg^{2+}$ -bound forms of 1 in EtOH–H<sub>2</sub>O (1:1, v/v) and in water solutions are shown in Fig. S4 and S5 (ESI†). From pH 2 to pH 10, absorption intensities are essentially insensitive to pH. Such photophysical properties of **1** in the presence and in the absence of  $Hg^{2+}$  suggest that this sensor may be very useful in practical applications.

In summary, we have reported here a simple yet highly selective colorimetric fluoroionophore (chemosensor **1**) for aqueous  $Hg^{2+}$ based on ICT in which the metal ionophore has been incorporated into the electron donor moiety of the fluorophore. The chemosensor shows a remarkably high ability to discriminate between  $Hg^{2+}$  and chemically similar ions in conjunction with a visible colorimetric change from red to colorless, leading to both "naked-eye" and fluorometric detection of  $Hg^{2+}$  cations.

### **Acknowledgements**

This project was partially sponsored by NCET-09-0894, the National Natural Science Foundation of China (NO: 20972111, 21074093, 21004044), SRF for ROCS, SEM, the Natural Science Foundation of Tianjin (NO: 08JCYBJC26700) and the State Key Lab. Elemental-Organic Chemistry at Nankai University (NO.0908 and 1011).

### **Notes and references**

- 1 E. M. Nolan and S. J. Lippard, *Chem. Rev.*, 2008, **108**, 3443; S. Lee, I. Choi, S. Hong, Y. I. Yang, J. Lee, T. Kang and J. Yi, *Chem. Commun.*, 2009, 6171.
- 2 (*a*) A. Renzoni, F. Zino and E. Franchi, *Environ. Res.*, 1998, **77**, 68; (*b*) H. H. Harris, I. J. Pickering and G. N. George, *Science*, 2003, **301**, 1203; (*c*) J. M. Benoit, W. F. Fitzgerald and A. W. Damman, *Environ. Res.*, 1998, **78**, 118; (*d*) D. W. Boening, *Chemosphere*, 2000, **40**, 1335.
- 3 (*a*) T. W. Clarkson, L. Magos and G. J. N. Myers, *N. Engl. J. Med.*, 2003, **349**, 1731; (*b*) B. R. Von, *J. Appl. Toxicol.*, 1995, **15**, 483.
- 4 (*a*) N. Basu, A. Scheuhammer, N. Grochowina, K. Klenavic, F. Evans, M. Obrien and M. Chan, *Environ. Sci. Technol.*, 2005, **39**, 3585; (*b*) P. Grandjean, R. F. White and F. Debes, *Environ. Res.*, 1998, **77**, 165.
- 5 Selective chemosensors based on PET: (*a*) E. M. Nolan and S. J. Lippard, *J. Am. Chem. Soc.*, 2003, **125**, 14270; (*b*) X. Guo, X. Qian and L. Jia, *J. Am. Chem. Soc.*, 2004, **126**, 2272; (*c*) L. Prodi, C. Bargossi, M. Montalti, N. Zaccheroni, N. Su, J. S. Bradshaw, R. M. Izatt and P. B. Savage, *J. Am. Chem. Soc.*, 2000, **122**, 6769. Selective chemosensors

based on ICT: (*d*) J. V. Ros-Lis, R. Martinez-Màñez, K. Rurack, F. Sancenon, J. Soto and M. Spieles, *Inorg. Chem.*, 2004, 43, 5183; (*e*) J. Wang and X. Qian, *Chem. Commun.*, 2006, 109; (*f*) O. Brümmer, J. J. La Clair and K. D. Janda, *Org. Lett.*, 1999, **1**, 415; (*g*) M. J. Choi, M. Y. Kim and S. K. Chang, *Chem. Commun.*, 2001, 1664; (*h*) K. Rurack, M. Kollmannsberger, U. Resch-Genger and J. Daub, *J. Am. Chem. Soc.*, 2000, **122**, 968; (*i*) H. Sakamoto, J. Ishikawa, S. Nakao and H. Wada, *Chem. Commun.*, 2000, 2395; (*j*) R. Martinez-Màñez, R. Radeglia, K. Rurack and J. Soto, *J. Am. Chem. Soc.*, 2003, **125**, 3418. Selective chemosensors based on FRET: (*k*) X. Zhang, Y. Xiao and X. Qian, *Angew. Chem., Int. Ed.*, 2008, **47**, 8025.

- 6 (a) F. Sancenón, R. Martinez-Màñez and J. Soto, *Chem. Commun.*, 2001, 2262; (*b*) M.-Y. Chae and A. W. Czarnik, *J. Am. Chem. Soc.*, 1992, 114, 9704; (c) J. V. Ros-Lis, M. D. Marcos, R. Martinez-Màñez, K. Rurack and J. Soto, *Angew. Chem., Int. Ed.*, 2005, **44**, 4405; (*d*) G. Hennrich, H. Sonnenschein and U. Resch-Genger, *J. Am. Chem. Soc.*, 1999, **121**, 5073; (*e*) M. G. Choi, D. H. Ryu, H. L. Jeon, S. Cha, J. Cho, H. H. Joo, K. S. Hong, C. Lee, S. Ahn and S.-K. Chang, *Org. Lett.*, 2008, **10**, 3717; (*f*) M. H. Lee, B.-K. Cho, J. Yoon and J. S. Kim, *Org. Lett.*, 2007, **9**, 4515; (*g*) K. C. Song, J. S. Kim, S. M. Park, K.-C. Chung, S. Ahn and S.-K. Chang, *Org. Lett.*, 2006, **8**, 3413; (*h*) H. Zheng, Z.-H. Qian, L. Xu, F.-F. Yuan, L.-D. Lan and J.-G. Xu, *Org. Lett.*, 2006, **8**, 859; (*i*) Y. Chung and K. H. Ahn, *J. Org. Chem.*, 2006, **71**, 9470; (*j*) S.-K. Ko, Y.-K. Yang, J. Tae and I. Shin, *J. Am. Chem. Soc.*, 2006, **128**, 14150; (*k*) Y. Zhao, Z. Lin, C. He, H. Wu and C. Duan, *Inorg. Chem.*, 2006, **45**, 10013; (*l*) O. del Campo, A. Carbayo, J. V. Cuevas, A. Munoz, G. Garcıa-Herbosa, D. Moreno, E. Ballesteros, S. Basurto, T. Gomez and T. Torroba, *Chem. Commun.*, 2008, 4576; (*m*) W. Jiang and W. Wang, *Chem. Commun.*, 2009, 3913; (*n*) X.-J. Zhu, S.-T. Fu, W.-K. Wong, J.-P. Guo and W.-Y. Wong, *Angew. Chem., Int. Ed.*, 2006, **45**, 3150.
- 7 S. A. Malashikhin, K. K. Baldridge and N. S. Finney, *Org. Lett.*, 2010, **12**, 940.
- 8 (*a*) A. Coskun and E. U. Akkaya, *J. Am. Chem. Soc.*, 2006, **128**, 14474; (b) F. Sancenón, R. Martínez-Màñez and J. Soto, *Chem. Commun.*, 2001, 2262; (*c*) Y.-K. Yang, K.-J. Yook and J. Tae, *J. Am. Chem. Soc.*, 2005, **127**, 16760; (*d*) E. M. Nolan, M. E. Racine and S. J. Lippard, *Inorg. Chem.*, 2006, 45, 2742; (*e*) A. Caballero, R. Martínez, V. Lloveras, I. Ratera, J. Vidal-Gancedo, K. Wurst, A. Tárraga, P. Molina and J. Veciana, *J. Am. Chem. Soc.*, 2005, **127**, 15666; (*f*) Y. Zhao and Z. Zhong, *J. Am. Chem. Soc.*, 2006, **128**, 9988; (*g*) A. Caballero, R. Mart´ınez, V. Lloveras, I. Ratera, J. Vidal-Gancedo, K. Wurst, A. Tárraga, P. Molina and J. Veciana, *J. Am. Chem. Soc.*, 2005, **127**, 15666.
- 9 (*a*) F. Song, S. Watanabe, P. E. Floreancig and K. Koide, *J. Am. Chem. Soc.*, 2008, **130**, 16460; (*b*) X. Guo, X. Qian and L. Jia, *J. Am. Chem. Soc.*, 2004, **126**, 2272; (*c*) A. B. Descalzo, R. Mart´ınez-Ma`nez, R. ˜ Radeglia, K. Rurack and J. Soto, *J. Am. Chem. Soc.*, 2003, **125**, 3418; (*d*) E. M. Nolan and S. J. Lippard, *J. Am. Chem. Soc.*, 2007, **129**, 5910; (*e*) S. Yoon, A. E. Albers, A. P. Wong and C. J. Chang, *J. Am. Chem. Soc.*, 2005, **127**, 16030; (*f*) J. Wang and X. Qian, *Chem. Commun.*, 2006, 109; (g) K. G. Vaswani and M. D. Keränen, *Inorg. Chem.*, 2009, 48, 5797; (*h*) B. Tang, B. Ding, K. Xu and L. Tong, *Chem.–Eur. J.*, 2009, **15**, 3147; (*i*) W. Jiang and W. Wang, *Chem. Commun.*, 2009, 3913.
- 10 (*a*) A. Mishra, R. K. Behera, P. K. Behera, B. K. Mishra and G. B. Behera, *Chem. Rev.*, 2000, **100**, 1973; (*b*) S. Tatay, P. Gavia, E. Coronado and E. Palomares, *Org. Lett.*, 2006, **8**, 3857.
- 11 M. Boiocchi, D. L. Boca, E. D. Gomez, L. Fabbrizzi, M. Licchelli and ´ E. Monzani, *J. Am. Chem. Soc.*, 2004, **126**, 16507.
- 12 (*a*) Y. Che, X. Yang and L. Zang, *Chem. Commun.*, 2008, 1413; (*b*) Standardization Administration (SA) of the People's Republic of China, Integrated Wastewater Discharge Standard, GB 8978, 1996; (*c*) Regulatory Impact Analysis of the Clean Air Mercury Rule, US EPA, Research Triangle Park, NC, 2005, EPA-452/R-05 003.